Independent variation in photosynthetic capacity and stomatal conductance leads to differences in intrinsic water use efficiency in 11 soybean genotypes before and during mild drought

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Abstract

Intrinsic water use efficiency (WUEintr), the ratio of photosynthesis to stomatal conductance to water, is often used as an index for crop water use in breeding projects. However, WUEintr conflates variation in these two processes, and thus may be less useful as a selection trait than knowledge of both components. The goal of the present study was to determine whether the contribution of photosynthetic capacity and stomatal conductance to WUEintr varied independently between soybean genotypes and whether this pattern was interactive with mild drought. Photosynthetic capacity was defined as the variation in WUEintr that would occur if genotypes of interest had the same stomatal conductance as a reference genotype and only differed in photosynthesis; similarly, the contribution of stomatal conductance to WUEintr was calculated assuming a constant photosynthetic capacity across genotypes. Genotypic differences in stomatal conductance had the greatest effect on WUEintr (26% variation when well watered), and was uncorrelated with the effect of photosynthetic capacity on WUEintr. Thus, photosynthetic advantages of 8.3% were maintained under drought. The maximal rate of Rubisco carboxylation, generally the limiting photosynthetic process for soybeans, was correlated with photosynthetic capacity. As this trait was not interactive with leaf temperature, and photosynthetic capacity differences were maintained under mild drought, the observed patterns of photosynthetic advantage for particular genotypes are likely to be consistent across a range of environmental conditions. This suggests that it is possible to employ a selection strategy of breeding water-saving soybeans with high photosynthetic capacities to compensate for otherwise reduced photosynthesis in genotypes with lower stomatal conductance.

Key words: Breeding strategies, drought stress, intrinsic water use efficiency, photosynthesis, soybean, stomatal conductance.

Introduction

Every year US soybean [Glycine max (L.) Merr.] fields transpire more water than that which evaporates from all the Great Lakes combined (Snyder, 1960; Purcell et al., 2007). Given that irrigation and rainfall limitations on agriculture are expected to grow in the coming decades (Wang, 2005), the development of soybean genotypes with enhanced drought tolerance, escape or avoidance traits would be highly desirable. To date considerable genetic variation in soybean whole-plant transpiration (Fletcher et al., 2007; Sinclair et al., 2008b; Sadok and Sinclair, 2009a) and whole-plant water use efficiency (WUEwholeplant) has been demonstrated (Hufstetler et al., 2007). A particularly promising trait is the break in the
linear relationship between the whole-plant transpiration rate (TR), measured under well-watered and light-saturating conditions, and atmospheric vapour pressure deficit (VPD) that has been demonstrated for some soybean genotypes (Sadok and Sinclair, 2009a). These genotypes show a linear TR to VPD response under mesic conditions but have constant whole-plant TR at higher (drier) VPDs, putatively related to leaf-level hydraulic limitations (Sinclair et al., 2005, 2010). This is important as a large proportion of US soybeans are grown under water-limiting conditions (Boyer, 1982), and simulations suggest that in 70% of years many soybean-growing areas would benefit from a constant TR at high VPD (Sinclair et al., 2010). Similarly, an early reduction in stomatal conductance upon low soil water contents would be beneficial to productivity in many areas of the USA (Sinclair et al., 2010).

One unstudied effect is whether genotypes with traits such as constant TR at high VPD, or reduced stomatal conductance at low soil water content, also have depressed productivity through low stomatal conductance and thus lower photosynthetic rates. This would seem a severe penalty for breeding water-conserving plants. However, variation in photosynthesis is also dependent on the ability of a leaf to draw down CO₂ within the leaf—defined here as photosynthetic capacity. If photosynthetic capacity is independent of stomatal conductance to water, then breeding for increased photosynthetic capacity offers a way around this trade-off.

Intrinsic water use efficiency (WUEintr), the leaf-level ratio of photosynthesis to stomatal conductance (an indication of leaf transpiration), has been widely used to screen for heritable genotypic variation in water use characteristics of crops, as has the whole-plant equivalent (WUEwholeplant). As both the drawdown of CO₂ by Rubisco and diffusion through the stomata result in fractionation of carbon isotopes, a mass spectrometer can measure WUEintr instantaneously or in-integrated for the lifespan of the CO₂ fixed into cellular components in a tissue (Seibt et al., 2008). However, if the measurements of carbon isotope discrimination are not combined with knowledge of the variation in the components of WUEintr, then it is difficult to draw conclusions about whether WUEintr changes might be beneficial to crop water use or productivity, or both (see Theory section). That is, a crop with low photosynthetic capacity and stomatal conductance to water (the components of WUEintr) may have the same WUEintr as a crop with proportionally higher photosynthetic capacity and stomatal conductance to water. Indeed for soybeans, carbon isotope discrimination, and, by extension WUEintr, are not strongly related to WUEwholeplant (Specht et al., 2001). In contrast, carbon isotope fractionation in combination with gas exchange measurements has been useful for breeding drought-adapted wheat genotypes (Condon et al., 1990, 2002), and such gas exchange measurements, accounting for the components of WUEintr, have been applied to other crops (Petitgrew and Turley, 1998; Centritto et al., 2009).

However, it is felt that a simple experimental approach that would allow for determination of WUEintr components is missing.

Here practical formulations that enable the two components of WUEintr to be measured are defined. This allows testing of: (i) whether there is a trade-off between reduced transpiration in ‘constant TR’ genotypes and photosynthesis at high VPDs; (ii) whether there are photosynthetic differences between genotypes, independent of variation in gH₂O, and whether these are maintained under conditions necessary for this trait to be useful in the field such as mild drought or varying temperature conditions; and (iii) which traits determine the photosynthetic differences. The answer to these questions then enables a conceptual framework to be offered by which water-saving soybean genotypes may be bred.

**Theory**

Water use efficiency of a crop is defined as a yield, or carbon gain, relative to water used to fix that carbon (or grain). Much of the exchange of water and CO₂ is determined at the leaf level, where stomata co-regulate the influx of CO₂ to photosynthesis and the transpirational loss of water to the atmosphere. As transpiration is determined by both atmospheric dryness (VPD) and stomatal characteristics it does not relate directly to photosynthesis. For this reason, WUEintr is defined as the ratio of photosynthesis to stomatal conductance to water, thereby accounting for the effects of VPD on transpiration (Seibt et al., 2008). Similarly, photosynthetic rate (A) is determined by two processes:

\[ A = (g_{CO₂} \times (C_a - C_i)) \]

where \( g_{CO₂} \) is the stomatal conductance of CO₂ into the leaf and \( C_a - C_i \) is the CO₂ drawdown, due to photosynthesis, between the atmosphere \( (C_a) \) and the intercellular \( (C_i) \) CO₂ concentrations. Stomatal conductance to CO₂ is strictly proportional to stomatal conductance to water \( (g_{H₂O} = g_{H₂O}/1.6) \), while the drawdown in CO₂ is proportional to the amount of nitrogen invested in photosynthetic components of the leaf such as Rubisco (Evans, 1989; Wright et al., 2003).

The relationship between A and \( g_{H₂O} \) is not linear for many species including soybeans, therefore, WUEintr is affected by two independent sources of variation: (i) variation in the position along this relationship, relating to \( g_{CO₂} \) and \( g_{H₂O} \), and (ii) variation in the height of the relationship, relating to photosynthetic capacity or ability to drawdown CO₂ (Fig. 1). By comparing the typical operating points of two genotypes and knowing the relationship between A and \( g_{H₂O} \), the contributions of variation in photosynthetic capacity (PC) and stomatal conductance \( (g_{H₂O}) \) to WUEintr, which are defined as WUEPC and WUEH₂O, respectively, can be estimated. Thus WUEPC is defined as the WUEintr that would occur if a genotype of interest had the same \( g_{H₂O} \) as a reference genotype and only differed in photosynthetic capacity. With reference to the
As the high WUEPC of the genotype of interest is maintained under WUE interest relative to the reference genotype, which has been termed from an increase in photosynthetic capacity of the genotype of interest. A soybean genotype of interest has an increased WUEintr relative to the average value measured for a reference genotype (triangle). This difference is due to a combination of two shifts: (i) a decrease in gH2O resulting in an increase in WUEintr, which has been termed WUEgH2O (circle; WUEgH2O=24.6/0.395=62.3μmol CO2 mol⁻¹ H2O) and (ii) the genotypes have different A to gH,O relationships, resulting from an increase in photosynthetic capacity of the genotype of interest relative to the reference genotype, which has been termed WUEgC (square; WUEgC=29.1/0.498=58.4μmol CO2 mol⁻¹ H2O). As the high WUEgC of the genotype of interest is maintained under lower gH,O, photosynthetic capacity differences can compensate for variation in water use. All points along a line passing through the origin (e.g. the dashed line) have the same WUEintr. The A to gH,O relationship and variation in WUEPC and WUEgH2O represent realistic values for soybean genotypes.

Theoretical example shown in Fig. 1, the WUEPC of the genotype of interest is:

\[
\text{WUEPC} = \frac{A_{\text{of genotype of interest} \text{ at } g_{H,O} \text{ of reference genotype}}}{g_{H,O(\text{of reference genotype})}}
\]

Similarly, WUEgH2O is the WUEintr that would occur if the genotype of interest had the same photosynthetic capacity (A to gH,O relationship) as the reference genotype and only differed in gH,O. In the case of Fig. 1, the WUEgH2O is:

\[
\text{WUEgH2O} = \frac{A_{\text{of reference genotype at } g_{H,O} \text{ of genotype of interest}}}{g_{H,O(\text{of genotype of interest})}}
\]

The resultant WUEintr for a soybean genotype of interest is then the combination of the change in WUEgH2O and WUEPC from the reference genotype (WUEintr=AgH2O of the genotype of interest).

To evaluate these two components of WUEintr considerable data are required, specifically, gas exchange measurements of a genotype of interest are needed in combination with an established A to gH,O relationship for both the genotype of interest and a reference genotype. As VPD changes may not result in variation in gH,O for the genotypes with a linear TR response to VPD, variation in soil water content may be used instead to cause the variation in gH,O needed to establish the A to gH,O relationships. This approach is based upon the empirically testable assumption that the A to gH,O relationship differs significantly between genotypes, and that it is relatively invariable within a genotype, as may be the case for sun-leaves in the uniform canopies of crops, particularly nitrogen fixers such as soybean. Virgona and Farquhar (1996) propose a similar approach, review its limitations, and use it to determine the reasons for differences in water use efficiency amongst sunflower genotypes in response to nitrogen treatments.

Two chloroplast processes may limit photosynthesis, individually or jointly: the CO₂-limited rate of Rubisco carboxylation, indicated by Vcmax, and the rate of RuBP regeneration (Jmax), estimated from the CO₂-saturated photosynthetic rate. These processes are differentially affected by leaf temperature, and a leaf may transition between these two limitations due to changes in stomatal conductance (Hikosaka, 1997; Hikosaka et al., 1999). Thus, it is possible that photosynthetic advantages under humid mesic conditions will not translate to conditions of mild drought, high temperatures associated with increasing VPD, or where stomata are limited to photosynthesis and other stresses are yet to influence productivity. There is also genetic variability in the TR to VPD relationship (Bunce, 1984; Sadok and Sinclair, 2009a). If there are interactions between genotype and mild drought for both WUEgH2O and WUEPC, it would complicate breeding efforts to improve soybean water use. Therefore, WUEPC and WUEgH2O have been measured over a range of conditions for 11 soybean genotypes, with the aim to elucidate whether genotype by environment interactions are present.

Materials and methods

Experiments were conducted in the Harvard Greenhouse Facility between August and November 2009 at Harvard University, Cambridge, MA, USA (42°N, 71°W). The first two experiments and the drought experiment were used to screen as many genotypes as possible for classic gas exchange traits, while the third experiment involved imposition of a mild drought (Table 1). For all experiments, air temperatures were maintained between a daily maximum of 27°C and minimum of 17°C measured within a screened box. The natural light environment was supplemented with a combination of 500W metal halide and high-pressure sodium lamps, enhancing the natural photosynthetic photon flux density (PPFD) by 500–600μmol m⁻² s⁻¹ for 16h d⁻¹.

Plant culture

Seeds were germinated in seedling trays, and transplanted to 2.2l pots once the first leaves had started to develop. Pots were filled with Farfard 3B potting soil including Osmocote Plus slow release fertilizer as per the manufacturer’s instructions (15/9/12 N/P/K/Mg+trace elements: S, B, Cu, Fe, Mn, Mo, Zn; Scotts Company). The pots
were watered daily and fertilized weekly with a water-soluble fertilizer (21S/20 N/P/K+trace elements: B, Cu, Fe, Mn, Mo, Zn; Peters Excel All Purpose fertilizer, Scotts Company).

Seeds of 11 soybean genotypes were provided by T.E. Carter (USDA-ARS, Raleigh, NC, USA), and selected to represent genotypes with a variety of TRs at high VPDs and a variety of TR to VPD responses (Table 2). Four and seven genotypes were chosen to represent genotypes with increasing TR at high VPD and constant TR at high VPDs, respectively. Of these 11 genotypes, the genotypes with increasing TR at high VPD were reported to have the highest TR at a VPD of 3.5kPa similar to that which occurred in these experiments (Table 2). PI416937 has been demonstrated to be slow wilting (King et al., 2009), corresponding to a constant TR at high VPD (Fletcher et al., 2007), and consequently was crossed with the standard cultivar Benning. The progeny, ‘G00BP’-labelled recombinant inbred lines (RILs), displayed a range of TR to VPD responses in excess of that shown by the parents, which both had constant TR at high VPD (Sadok and Sinclair, 2009a). Five of these RILs, and the parents, were selected to represent the range of these responses, including one genotype with increasing TR at high VPD, G00BP-147. In addition, a standard reference cultivar (Biloxi), two recent commercial cultivars (N96-7031 and N01-11136), and a second plant introduction (PI419738) were also included. PI419738 was chosen as it had a high WUE wholeplant relative to PI418937 (Hufstetler et al., 2007), suggesting that variation in WUE mix may be present, and was slow wilting (King et al., 2009); however, this is likely to be due to a mechanism different from that in PI416937 as this genotype has increasing TR at high VPD (Fletcher et al., 2007). N96-7031 has similar WUE wholeplant to PI416937 (Hufstetler et al., 2007), but an increasing TR to VPD response (Sadok and Sinclair, 2009a). N01-11136 represents the most extreme genotype with increasing TR at high VPD, with the highest whole-plant TRs (Sadok and Sinclair, 2009a). As the majority of the selected genotypes were descended from PI416937, this was selected as the reference genotype.

Drought experiment

Once the seedlings reached the V3 stage of growth, ~20–30cm tall and with between two and three fully expanded trifoliolate leaves, the pots were randomly assigned to a control (well-watered) or drought treatment. Sample sizes were five control and six drought treatment plants per genotype. Initial measurements were made on 1 and 2 November on all plants prior to imposition of drought. Pots were submerged in water to achieve saturation and, after allowing 2h for excess water to drain, the pot water-holding capacity was determined. Subsequently pot weights were measured every day, within a 2h period, after sunset. Upon imposition of the drought treatment on 3 November, all pots were watered to a predetermined weight, calculated to result in an 85% soil water content (SWC) for control plants and a standard SWC for drought-stressed plants. In this way, drought could be imposed in a consistent and gradual manner for all plants, regardless of leaf area. Drought was imposed over 7d, during which the SWC of the drought-treated plants was returned to 85, 75, 65, 50, or 45% SWC each day at the time of weighing. At the end of the experiment, the leaves and stems were removed, and pots were dried for 2 weeks in a forced draught oven at 85°C, and SWC was determined as (fresh weight−dry weight)/(field capacity weight−dry weight)×100. The average canopy air temperatures and humidity measured simultaneously with the gas exchange measurements for the drought experiment, using a screened Vaisala probe, ranged from 29.5°C and 14.2% to 28.5% relative humidity, corresponding to a range in mid-day average VPD of 3.0–4.0kPa. All drought-stressed plants (SWC <50%) had leaf temperatures well below the reported >33–35°C leaf temperature threshold for thermal inhibition of soybean photosynthesis (Harley et al., 1985), eliminating the possibility of artefacts due to the reduced evaporative cooling of drought-stressed leaves. The SWCs chosen to represent mesic and mild drought conditions corresponded to average leaf water potentials of −0.62MPa and −1.05MPa, respectively (leaf water potentials

Table 1. Summary of the experiments

<table>
<thead>
<tr>
<th>Experiment 1</th>
<th>Measurements</th>
<th>Sowing date</th>
<th>Measurement period</th>
<th>Conditions</th>
</tr>
</thead>
<tbody>
<tr>
<td>Experiment 2</td>
<td>Vcmax and Jmax gas exchange measurements</td>
<td>22 August</td>
<td>16–23 September</td>
<td>Well watered</td>
</tr>
<tr>
<td>Drought experiment</td>
<td>Vcmax and Jmax gas exchange measurements</td>
<td>7 October</td>
<td>1–12 November</td>
<td>Drought imposed on half of the plants</td>
</tr>
</tbody>
</table>

Table 2. VPD response characteristics and origin of the 11 soybean genotypes for which the drought experiment was undertaken

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Origin</th>
<th>WUEwholeplant (g DM kg⁻¹ H₂O)⁻¹</th>
<th>TR predicted for 3.5 kPa (mg H₂O m⁻² s⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td>N01-11136</td>
<td>NTCPR94- 5157×N96-7031</td>
<td>62.0</td>
<td>76.7</td>
</tr>
<tr>
<td>PI471938 Plant introduction from Nepal</td>
<td></td>
<td>3.38</td>
<td>75.3</td>
</tr>
<tr>
<td>G00BP-147</td>
<td>PI416937×Benning</td>
<td>2.81</td>
<td>63.7</td>
</tr>
<tr>
<td>N96-7031</td>
<td>N7001×N90-7241</td>
<td>2.81</td>
<td>62.0</td>
</tr>
<tr>
<td>PI416937 Plant introduction from Japan</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Biloxi Historical cultivar</td>
<td></td>
<td>3.14</td>
<td>34.6</td>
</tr>
<tr>
<td>G00BP-60</td>
<td>PI416937×Benning</td>
<td>59.6</td>
<td></td>
</tr>
<tr>
<td>G00BP-160</td>
<td>PI416937×Benning</td>
<td>48.5</td>
<td></td>
</tr>
<tr>
<td>G00BP-50</td>
<td>PI416937×Benning</td>
<td>39.8</td>
<td></td>
</tr>
<tr>
<td>G00BP-75</td>
<td>PI416937×Benning</td>
<td>39.3</td>
<td></td>
</tr>
<tr>
<td>Benning Hutcheson×Coker 6738</td>
<td></td>
<td>3.14</td>
<td></td>
</tr>
</tbody>
</table>

\( ^a \text{Hufstetler et al. (2007) } \)

\( ^b \text{Descendant of PI416937.} \)
were measured on six genotypes on the final day of drought for both well-watered and drought-stressed plants; n=4–5 per treatment). The most severe leaf water potentials were between −1.2MPa and −1.4MPa, close to the minimum extractable soil water potential for soybean of approximately −1.2MPa (Brisson et al., 1993; Sinclair et al., 2008a). Leaf water potentials were measured with a Scholander-type pressure chamber after transpiring leaves were bagged, excised, and allowed to equilibrate for 10 min in a dark cool container.

Gas exchange measurements
Gas exchange for the drought experiment was measured using a LI-6400 photosynthesis system with a red-blue light source and 6 cm² cuvette. At the start of the drought experiment the youngest fully expanded primary leaflet (mostly second trifoliolate) was marked and subsequently used for all gas exchange measurements. To standardize the environmental conditions, an hour before measurement each plant was placed on the edge of the greenhouse bench, with the marked leaf oriented to receive full sunlight. Gas exchange measurements were made on the majority of plants on 1, 4, 6, and 8 November from 10:00 to 15:00 hr. As not all plants could be measured on a single day, the remaining third of the plants were measured on 2 and 7 November. Cuvette conditions were: PPFD, 2000 μmol m⁻² s⁻¹; flow rate, 500 μmol s⁻¹; sample CO₂, 390 μmol mol⁻¹; VPD, 2.3 ± 0.5 kPa; leaf temperature, 31.3 ± 1.6°C (mean ± SD); and the cuvette fan was set to fast. Leaf temperature was allowed to track ambient conditions; however, the cuvette was shaded, and block temperature cooled to 32°C when leaf temperatures were >33°C.

Well-watered plants from the drought experiment and the two earlier experiments were used to determine the response of photosynthetic traits to leaf temperature. A screening approach was used to allow many plants to be measured and CO₂ response curve traits related to WUEPC. Namely, the CO₂ and light-saturated rate of photosynthesis was used to estimate the maximal rate of RuBP regeneration (Jmax), and the initial slope of the CO₂ response curve was used to provide an estimate of the Rubisco carboxylation capacity (Vcmax) (see Supplementary Fig S1. available at JXB online). Leaf temperature was allowed to vary with ambient conditions, which resulted in a >10°C range across days. CO₂-saturated photosynthetic rates were measured on the second trifoliolate, or marked leaf of on average 16 plants per genotype. The 2 cm² LI-6400 fluorescence chamber was used for this experiment. Cuvette conditions were: PPFD, 2000 μmol m⁻² s⁻¹; flow rate, 250 μmol s⁻¹; sample CO₂, 190 μmol mol⁻¹; VPD, 1.6 ± 0.3 kPa (mean ± SD); leaf temperature, 22.5–32.9°C. Additional full CO₂ response curves demonstrated that photosynthetic rates at high CO₂ were not significantly triose phosphate limited, thus photosynthetic rates at very high CO₂, where photorespiration is competitively inhibited, are approximately equal to a quarter of the maximal rate of RuBP regeneration (von Caemmerer, 2000) (Jmax ≈ 4 × A at 1990 μmol mol⁻¹ CO₂). Vcmax—the initial slope of the CO₂ response curve—was measured for a subset of the 11 genotypes by varying sample CO₂ from 225, 175, 125, 75, to 50 μmol mol⁻¹. The initial slope value was estimated from linear regression of points with CO₂ <200 μmol mol⁻¹, and was equated to the derivative of the Rubisco-limited photosynthesis function (von Caemmerer, 2000) at the photocompensation point, which was then solved for Vcmax using the temperature corrections and values of Kc, Kc, and Γ of von Caemmerer (2000). The genotypes used represented three with constant TR at high VPD (PI416937, G00BP-160, and Benning) and three with increasing TR at high VPD (N96-7031, PI471938, and N01-11136). The LI-6400 6 cm² cuvette was used for these experiments. Cuvette conditions were: PPFD, 2000 μmol m⁻² s⁻¹; flow rate, 400 μmol s⁻¹; VPD, 1.6 ± 0.6 kPa (mean ± SD); leaf temperature, 21.9–34.4°C. This data set was augmented with measurements of Vcmax made on 3–5 leaves of three other genotypes at leaf temperatures within 1°C of 30°C. Both sets of measurements were corrected for CO₂ leaks using an empirical relationship, where CO₂ leaks were measured during CO₂ response curves as the CO₂ concentration difference between the previously matched reference and sample infrared gas analysers (IRGAs) in an empty chamber with the above conditions.

Data analysis
The R statistical and graphical package (R Development Core Team, 2010) was used to perform general linear model (GLM) analyses as follows. As the photosynthesis versus stomatal conductance relationship scaled non-linearly, it was not appropriate to analyse WUEme as a ratio (Jasienski and Bazzaz, 1999), therefore a standard GLM approach to data analysis was adopted (Müller et al., 2000; Crawley, 2005). Hence, photosynthesis was analysed through a GLM of the natural logarithm of stomatal conductance to H₂O; similarly Jmax was predicted by leaf temperature, and the natural logarithm of Vcmax predicted by the natural logarithm of leaf temperature (a power function). In these analyses, genotype and the interaction with the linear predictor were initially included—testing for differences in slope. However, the presented results represent minimum adequate models where unnecessary (non-significant) higher order interactions have been removed using the step/ function of the R statistical package (Crawley, 2005). That is, if there was no significant genotype with linear predictor interaction, the slopes were assumed to be the same and this term was removed from the analysis, as per a standard analysis of covariance (ANCOVA). PI416937, an ancestral genotype of the majority of the genotypes measured in this study, was used as the reference genotype with which changes in most variables including WUEintr, WUEPC, and WUEgH₂O were compared, and relative to which the significance of linear model coefficients were compared. This avoided the use of many post hoc comparisons, whilst still providing information on significant differences between genotypes.

Stomatal conductance to H₂O was non-linearly related to SWC, with gH₂O values at high and low SWC reduced relative to SWCs at ~85%. A quadratic equation best accounted for this variation, and allowed a linear model analysis to be performed testing for effects of genotype, SWC, SWC², and interactions. To summarize these data, the SWCs used to regulate pot water content were selected to represent mesic (85% SWC) and drought conditions (50% SWC). Using the linear model for gH₂O to SWC, average values of gH₂O were estimated for each genotype and SWC. In turn, the average A value for these gH₂O values was estimated from the linear model for A to gH₂O. Using these averages and regression equations, values of WUEintr, WUEPC, and WUEgH₂O for each genotype could be estimated relative to PI416937.

Results
Intrinsic water use efficiency and variation in components
The relationship between A and gH₂O measured over the course of the mild drought had significant curvature, and the shape was highly conserved across the 11 soybean genotypes (Fig. 2; Supplementary Table S1. at JXB online), although there were differences between genotypes in the elevation of the curve. The genotype × gH₂O interaction was not significant (F10,358 = 0.703, P = 0.721), although genotype had a significant effect (Supplementary Table S1.). As this analysis used the natural logarithm of gH₂O to predict A, this signifies that the between-genotype differences in A for a given gH₂O were consistent across the range of gH₂O induced by drought. Thus, all genotypes apart from Biloxi and G00BP-59 had significantly higher A than PI416937 across the entire range of gH₂O (Supplementary Table S1.). As Biloxi, a historical
reference cultivar, was similar to PI416937 in both \( g_{H_{2}O} \) and \( A \), it appears that the choice of a plant introduction as reference genotype affects these results little.

WUE\text{intr} varied greatly between genotypes (Figs 3, 4), largely due to the curvature of the relationship between \( A \) and \( g_{H_{2}O} \) and points for all genotypes combined (light squares). The fitted relationships (using the natural logarithm of \( g_{H_{2}O} \)) are shown for each genotype (solid line) and relative to the relationship for PI416937 (dashed line). The final panel shows variation among genotypes in photosynthetic rate at the average \( g_{H_{2}O} \) of PI416937 at a SWC of 85% (0.498 mol m\(^{-2}\) s\(^{-1}\)). Statistical differences and standard errors of the mean are derived from Supplementary Table S1. at JXB online. Genotypes were ranked in order of greatest (N01-11136) to lowest \( g_{H_{2}O} \) (G00BP-59). Soybean genotypes with a constant TR at high VPD relationship (circles and solid bars) and genotypes with increasing TR at high VPD (triangles and hatched bars) are shown.

**Fig. 2** Differences between genotypes in photosynthesis (\( A \)) and intrinsic water use efficiency (WUE\text{intr} = \( A/g_{H_{2}O} \)) during drought-induced variation in stomatal conductance to \( H_{2}O (g_{H_{2}O}) \). Each panel shows measurements made for a soybean genotype (dark symbols) and points for all genotypes combined (light squares). The fitted relationships (using the natural logarithm of \( g_{H_{2}O} \)) are shown for each genotype (solid line) and relative to the relationship for PI416937 (dashed line). The final panel shows variation among genotypes in photosynthetic rate at the average \( g_{H_{2}O} \) of PI416937 at a SWC of 85% (0.498 mol m\(^{-2}\) s\(^{-1}\)). Statistical differences and standard errors of the mean are derived from Supplementary Table S1. at JXB online. Genotypes were ranked in order of greatest (N01-11136) to lowest \( g_{H_{2}O} \) (G00BP-59). Soybean genotypes with a constant TR at high VPD relationship (circles and solid bars) and genotypes with increasing TR at high VPD (triangles and hatched bars) are shown.

WUE\text{intr} varied greatly between genotypes (Figs 3, 4), largely due to the curvature of the relationship between \( A \) and \( g_{H_{2}O} \) and variation in \( g_{H_{2}O} \) between genotypes (Fig. 3; Supplementary Table S1.). The average genotype \( g_{H_{2}O} \) under well-watered conditions varied by 43%, between a high of 0.632 mol m\(^{-2}\) s\(^{-1}\) for N01-11136 and a low of 0.42 mol m\(^{-2}\) s\(^{-1}\) for G00BP-59, relative to the reference genotype, leading to 15% variation in \( A \), given no differences in photosynthetic capacity (Fig. 3). However, enhanced photosynthetic capacity resulted in up to 8.3% increases in \( A \) for N96-7031, 5.1% increases in G00BP-78, and 4.1% in Benning relative to PI416937. This resulted in increased WUE\text{intr} for genotypes with low \( g_{H_{2}O} \) [e.g. region (3)] or intermediate to high \( g_{H_{2}O} \) [e.g. region (2)], and in some genotypes the increased photosynthetic capacity entirely compensated for reductions in \( A \) due to low \( g_{H_{2}O} \) (e.g. Benning) (Fig. 3).

When shown as the percentage difference in WUE between a genotype of interest and the reference PI416937, WUE\text{intr} changed largely due to variation in WUE\text{g}\text{H_{2}O}, and to a lesser
degree with WUEPC (Fig. 4A). A quadratic function was used to conduct a linear model analysis of the relationship of \( g_{H,O} \) to SWC (Supplementary Table S1. at JXB online). The genotype by SWC interaction was significant, as was the SWC\(^2\) term. Due to the significant genotype by SWC interaction, the average \( g_{H,O} \) measured for genotypes under mesic conditions was uncorrelated to those measured under mild drought (\( R^2 = 0.254, n = 11, P = 0.114 \)), for average \( g_{H,O} \) of the genotypes at 50% and 85% SWC). Genotype \( g_{H,O} \) response interacted with SWC (Supplementary Table S1), and therefore WUE\(_{\text{intr}} \) advantages under mesic conditions were not maintained for all genotypes under drought conditions, although the WUEPC advantages were constant (Fig. 4B). Exceptions were Benning, which had high WUE\(_{\text{intr}} \) due to favourable WUE\(_{g_{H,O}} \) and WUEPC under both mesic and drought conditions, and N01-11136 which had low WUE\(_{\text{intr}} \) due to extremely high \( g_{H,O} \) under all conditions.

Mechanisms of variation in WUEPC

Knowledge of WUEPC seems particularly useful for breeders, because variation in photosynthetic capacity was not correlated to \( g_{H,O} \) (Figs 3, 4), and WUEPC did not interact with drought. It was thus sought to establish the mechanisms underlying variation in WUEPC, and to establish a link with a trait that would allow easy field selection of high WUEPC genotypes. Leaf mass area (LMA), a rough estimate of the resource investment in a unit area of leaf (Wright et al., 2004), did not predict WUEPC (Fig. 5A), although there was significant variation between genotypes (\( F_{10,103} = 4.08, P < 0.002 \)), \( J_{\text{max}} \) which limits photosynthesis at low light or high CO\(_2\), corresponded only roughly to WUEPC (Fig. 5B), though there were significant differences between genotypes in \( J_{\text{max}} \) (Supplementary Table S2 at JXB online). \( V_{\text{c,max}} \), which typically limits photosynthesis at low to intermediate \( g_{H,O} \), correlated significantly to WUEPC for six genotypes for which temperature responses were measured (Fig. 5C); the inclusion of three extra genotypes measured at a set temperature (unfilled symbols) confirmed the relationship. Across an \( \sim 13^\circ \)C range of leaf temperatures the genotypes displayed a temperature effect for \( V_{\text{c,max}} \) and \( J_{\text{max}} \) significantly affecting the elevation of the \( V_{\text{c,max}} \) or \( J_{\text{max}} \) to \( T_{\text{leaf}} \) relationship, but not the slope (Supplementary Fig S2., Fig S3.). Thus, the differences in \( V_{\text{c,max}} \) and \( J_{\text{max}} \) across genotypes were consistent across all \( T_{\text{leaf}} \) conditions. Consistent with this pattern, \( T_{\text{leaf}} \) did not have a significant effect on WUE\(_{\text{intr}} \) (\( F_{1,357} = 0.428, P > 0.513 \)), or interact with genotype (\( F_{10,357} = 0.634, P > 0.785 \) for the model WUE\(_{\text{intr}} \) genotype \( \times T_{\text{leaf}} + \text{SWC} + \text{error} \)). Modelling of the measured \( V_{\text{c,max}} \) and \( J_{\text{max}} \) values using standard photosynthetic functions (von Caemmerer, 2000) demonstrated that \( V_{\text{c,max}} \) was limiting, or co-limiting, to photosynthesis at virtually all values of \( g_{H,O} \) (data not shown). The average \( Q_{10} \) values for all genotypes combined over the measured 22–32°C range were 2.15 and 1.67 for \( V_{\text{c,max}} \) and \( J_{\text{max}} \), respectively. As these compare favourably with published values, which indicate a higher \( Q_{10} \) value for \( V_{\text{c,max}} \) than \( J_{\text{max}} \) e.g. \( Q_{10} = 2.21 \) and 1.65 from 25°C to 35°C (von Caemmerer, 2000), the fitting procedures adopted here appear adequate. Thus, genotypes with increased \( V_{\text{c,max}} \) consistently led to WUEPC advantages during mesic and drought conditions and over a 10°C range of temperatures.

Discussion

Breeding strategies relating to soybean water use

A plant could be bred to avoid drought through conservation of soil moisture by reducing stomatal conductance to water at high VPD (a constant TR at high VPD) and thus avoiding future drought. While this strategy would result in a net increase in WUE\(_{\text{intr}} \) (Fig. 1), it would be at the expense of photosynthesis during periods of high VPD. Typical stomatal conductance reductions of 33% would result in 14% reductions in photosynthesis under high VPD, well-watered conditions, and would be larger in the field (personal observation) where hotter temperatures, wind, and soil water depletion are more important. Therefore, it would be beneficial to breed such water-conserving plants with enhanced WUEPC to compensate for the photosynthetic loss

Fig. 3 Variation in the average photosynthesis (A) and stomatal conductance to H\(_2\)O (\( g_{H,O} \)) operating points between well-watered soybean genotypes relative to the fitted A to \( g_{H,O} \) relationship for PI416937 (solid line) and N96-7031 data (dotted line). Soybean genotypes with a constant TR at high VPD (circles) and genotypes with increasing TR at high VPD (triangles) are shown. The diagonal dashed line represents points of equal intrinsic water use efficiency to PI416937. The numbers and shaded regions represent four categories of soybean genotypes having: (1) increased photosynthetic rates and reduced \( g_{H,O} \) relative to PI416937; (2) higher photosynthesis, but increased \( g_{H,O} \) relative to PI416937—genotypes have a range of WUE\(_{\text{intr}} \), some greater or less than PI416937; (3) reduced A and \( g_{H,O} \) relative to PI416937; and (4) generally less WUE\(_{\text{intr}} \) than PI416937. These categories are described in detail in the Discussion.
associated with reduced stomatal conductance at high VPD. Breeding plants with low water use during mild drought could also involve breeding for an increase in photosynthetic capacity (WUEPC), thus enabling the plant to photosynthesize under stomatal limiting conditions, where turgor-related growth inhibition may be present but water stress effects are not severe (Hsiao, 1973). This assumes that a photosynthetic capacity advantage is maintained under such drought conditions. These two strategies of water conservation and mild drought tolerance are not mutually exclusive as both could involve selection for increased photosynthetic capacity.

Fortunately, it appears that for soybean genotypes photosynthetic capacity is a constitutive trait, which does not interact with drought or temperature and is thus a potentially useful characteristic for breeders. However, the rankings of genotypes relative to TR do vary with VPD (Sadok and Sinclair, 2009a). This combination of differentially varying traits offers much choice for breeders. As photosynthetic capacity is independent of the presence of the constant TR at high VPD trait, this signifies that reduced productivity is not necessarily a consequence of breeding for water conservation under high VPD, and that the selection strategies suggested here may be successful.

Three possible breeding strategies present themselves when choosing a genotype of interest for breeding relative to a reference genotype, and based upon gas exchange measurements made under well-watered conditions. First, one could select for high photosynthetic rates (which it is assumed translate into some productivity advantage) through either increased $g_{H2O}$ or photosynthetic capacity. It is important to note that this strategy corresponds to the plants in region (2) in Fig. 3, and can be consistent with increases or decreases in $WUE_{intr}$. It seems likely that such plants, while using the same amount as (e.g. N96-7031 and G00BP-78) or more water than (e.g. N01-11136) the reference genotype (PI416937) or the standard cultivar Biloxi would have the potential for higher productivity and thus be generally desirable for non-water-limiting conditions. While it is difficult to extrapolate from greenhouse data as to which genotypes would be useful in the field, it does seem evident that genotypes with increasing TR at high VPD exemplify this selection strategy (Fig. 3). Past work suggests that this strategy was employed for Canadian short season soybeans over the last half-century (Morrison et al., 1999), although it is not necessarily the case that photosynthetic increases will increase productivity (Ford et al., 1983).

The second selection strategy is to breed plants that conserve water but also maintain some photosynthetic capacity advantage relative to the reference genotype [selection of genotypes in region (3) of Fig. 3]. Although plants
selected for these traits may avoid drought through water conservation during mesic periods, this would be associated with a reduction in productivity as there is only a partial photosynthetic compensation through higher photosynthetic capacities. The third selection strategy involves breeding genotypes for region (1) of Fig. 3, where reduced water use is associated with potential for higher productivity through higher photosynthetic capacities. While no soybean genotypes tested in this study filled this space, one may envisage a progeny line from a cross of N96-7031 and Benning, with a combination of the low stomatal conductance of Benning and the high photosynthetic capacity of both parents falling into region (1). Indeed, the observed 8.3% increase in photosynthetic capacity for N96-7031 would compensate for photosynthetic decreases caused by a constitutive 20% decrease in \( g_{H,O} \). Genotypes in region (4) would offer little benefits to breeders, as they would have reduced photosynthetic rates relative to the reference genotype’s \( A \) to \( g_{H,O} \) relationship. Note that some of the areas within region (4)—areas above the dashed line, but below the curve—actually have greater WUE\(_{intr}\) than PI416937, but would have depressed photosynthetic rates relative to a genotype with equivalent photosynthetic capacity to PI416937, and reduced \( g_{H,O} \). This illustrates the point that increased WUE\(_{intr}\) relative to the reference genotype is not necessarily associated with advantageous changes in the components of WUE\(_{intr}\).

These considerations are also valid at a whole-plant level: a high WUE\(_{whole\,plant}\) genotype is only of interest if it has a high productivity (Sinclair and Muchow, 2001), a fact not immediately obvious from the value of WUE\(_{whole\,plant}\). The breeding strategies suggested here are obviously contingent upon a number of assumptions. Do leaf-level processes directly scale with crop-level productivity and water use? Are photosynthetic advantages under well-watered conditions representative of drought or varying environmental conditions?

In practice, variation in transpiration, and \( g_{H,O} \), is likely to have a greater effect on WUE\(_{intr}\) than photosynthetic capacity when comparing genotypes within a crop species (Blum, 2005, 2009), and measured under controlled conditions (no variation in nitrogen or light). This is due to the large curvature of the \( A \) to \( g_{H,O} \) response for soybean, and the typical high position of photosynthetic operating points on this response in well-watered crops (Fig. 2). This curvature results in seemingly beneficial increasing WUE\(_{intr}\) under mild drought or reduced \( g_{H,O} \) at high VPD (Fig. 2), but it is important to note that it is also associated with decreasing \( A \). A number of other studies confirm that WUE\(_{whole\,plant}\) can be increased under mild drought (Hubick and Farquhar, 1989), while others find little change (Hubick et al., 1986; Liu and Stutzel, 2004). This variation may relate to the position of the typical operating point of these species on the \( A \) to \( g_{H,O} \) Curve. Obviously, reductions in leaf-level transpiration are concomitant with mild drought-induced productivity decreases, and therefore undesirable. Thus it is important for mild-drought stressed—\( g_{H,O} \)-limited—plants, or genotypes with constitutive traits conferring low \( g_{H,O} \), to have increased photosynthetic capacities to compensate for this effect.

Fig. 5 Relationships of photosynthetic capacity, estimated as WUE at a common \( g_{H,O} \) (WUE\(_{PC}\)), to average leaf mass area (A), the rate of RuBP regeneration (\( J_{max} \)) (B), and the maximal rate of Rubisco carboxylation (\( V_{c,max} \)) (C). \( J_{max} \) and \( V_{c,max} \) values were corrected to a common leaf temperature of 31.3°C (the average leaf temperature at which WUE\(_{PC}\) was measured), using fitted linear or power temperature response functions. WUE\(_{PC}\) was calculated by using photosynthetic values derived from Fig. 3 assuming the same stomatal conductance to H\(_2\)O (0.498mol m\(^{-2}\) s\(^{-1}\), the average value for PI416937 at 85% SWC). Open symbols in (C) represent average data for leaves measured within 1°C of 30°C (n >3), and temperature corrected using the fitted power response function used for the other genotypes. Soybean genotypes with a constant TR at high VPD (circles) and genotypes with increasing TR at high VPD (triangles) are shown.
The nature of variation in photosynthetic capacity

During the transition from mesic conditions to those of high VPD or mild drought, stomatal conductance is likely to become increasingly limiting to the supply of CO₂ for photosynthesis. The major biochemical limitations to photosynthesis, V_cmax and J_max, are less likely to change directly with VPD although between-genotype variation in these traits is a likely cause of differences in photosynthetic capacity. However, high VPD is dependent on high temperatures, which affect whether V_cmax or J_max is limiting to photosynthesis (Hikosaka, 1997; Hikosaka et al., 1999). Therefore, it is necessary to determine both the cause of the observed differences in photosynthetic capacity and the variation of photosynthetic capacity limitations with temperature. From the measured temperature response data it was possible to use standard photosynthetic models to determine which process was limiting to photosynthesis under given temperature and g_H₂O conditions (von Caemmerer, 2000). The majority of photosynthesis measurements were on the portion of the CO₂ response limited by V_cmax across the observed range of g_H₂O. Thus, the consistency of the photosynthetic advantages across a broad range of g_H₂O is partially due to the lack of limitation by J_max under mesic conditions that would be associated with transitions to V_cmax limitation under drought. Consistent with this finding is the correlation between WUE_P and V_cmax, but not J_max (Fig. 5). Thus, V_cmax appears to be the major source of variation in photosynthetic rates between soybean genotypes for a given g_H₂O. It is possible that the consistent V_cmax limitation is an artefact of the study being conducted in greenhouse conditions, resulting in greater leaf light capture capacity than may occur in the field. However, Rubisco-limited photosynthesis in the current study is consistent with Bernacchi et al. (2005) who found that field-grown soybeans were largely Rubisco or co-limited for much of the year, with increasing J_max over the growth season.

A second source of consistent photosynthetic capacity advantages may be due to the similarity of V_cmax and J_max temperature responses between genotypes. Any differences in the temperature responses of J_max and V_cmax would be important as they determine the co-limitation point of the CO₂ response curve, and therefore which factor limits photosynthesis. A third source of variation in A over a range of g_H₂O may be shifts from stomatal to metabolic limitations of photosynthesis under drought conditions (Medrano et al., 2002; Ennahli and Earl, 2005; Lawlor and Tezara, 2009). This did not appear to be the case for the plants measured during the drought (based upon photosynthetic modelling of observed C_i, A, V_cmax, and J_max; data not shown). The magnitude of the stomatal and metabolic limitations increased similarly with decreasing g_H₂O and did not appear to differ between genotypes. As differential metabolic limitations may dramatically change the shape of the A to g_H₂O relationship at low g_H₂O (Ripley et al., 2010), it is important that there are no systematic genotype differences. Metabolic limitations are likely to become important under severe drought—a state which the water-saving genotypes are more able to avoid.

Extension of these breeding strategies to the field

Inherent in these strategies are a number of assumptions concerning the scaling up of leaf-level processes to the canopy (Richards, 2000; Long et al., 2006): for instance, that the crop’s total transpiring leaf area remains unchanged during drought, or that the leaf is well coupled with environmental conditions (Sinclair et al., 1984). Thus, while knowledge of components of WUE_int may be a useful tool to identify potential breeding genotypes, whole-plant and field validations of how well these traits scale up to canopy conditions are vital.

These results are also predicated upon a positive relationship between photosynthesis and seed yield under breeding conditions. Past studies have shown varying correlations between photosynthesis and seed yield in soybean: with selection for A being unrelated to yield (Ford et al., 1983), and other studies demonstrate that canopy photosynthesis may relate to yield (Wells et al., 1982; Ashley and Boerma, 1989). While photosynthesis appears to be a heritable trait (Wiebold et al., 1981; Secor et al., 1982), the probable low potential for increasing leaf-level photosynthetic capacity may constrain the potential for differences in yield that one may find after generations of breeding for high A (Ashley and Boerma, 1989). However, the genotypes selected here showed sufficient genetic variability in photosynthetic capacity, through increases in V_cmax (Fig. 5), to allow some compensation for considerable reductions in g_H₂O, suggesting that there is some potential for breeding soybeans with higher photosynthetic rates. Further increases in photosynthetic rates and yields might be possible via a combination of selection for decreased resistance to CO₂ via increased stomatal conductance (Morrison et al., 1999), and possibly through alternative mechanisms such as increased mesophyll conductance to CO₂ (Barbour et al., 2010). Regardless, it seems that it is necessary to ask the opposite question: ‘Does reduced photosynthesis, associated with limited water use when TR is constant at high VPD or g_H₂O low under mild drought cause reductions in grain yield?’ If so, increases in photosynthetic capacity that compensate for stomatal limitations to photosynthesis may allow water-saving genotypes to retain the high yields of less water-conservative genotypes.

Field validation of this work is required before these breeding strategies are applied to the particular genotypes studied here. It is expected that for soybean the principle of photosynthetic capacity being non-interactive with mild drought would hold in field situations, particularly if V_cmax limits photosynthesis. These breeding strategies may not be entirely applicable to other crop species, as: (i) the shape of the A to g_H₂O relationship may be more linear, e.g. Panicum virgatum and Miscanthus x giganteus (Dohleman et al., 2009), resulting in a weak link between g_H₂O and WUE_int; (ii) photosynthesis may be limited by processes other than Rubisco carboxylation (e.g. RuBP regeneration), leading to interactions in photosynthetic capacity with drought or other environmental conditions; and (iii) increases in photosynthetic capacity may depend upon...
greater leaf nitrogen contents, a possible trade-off when these strategies are applied to non-nitrogen fixers or when there is pressure to reduce fertilizer inputs (Sinclair and Purcell, 2005).

Conclusion

WUE_{intr} can be conveniently measured as the ratio $A/g_{H_2O}$ or by carbon isotope discrimination. However, as has been shown here, WUE_{intr} is comprised of two components that vary independently of each other, one of which is interactive with drought. Thus we suggest that breeding soybean genotypes for drought tolerance, escape or avoidance requires more than an analysis of leaf-level WUE_{intr}. Instead, the components of WUE_{intr}: variation in photosynthetic capacity and stomatal conductance to H$_2$O, distinguish between the separate reasons for WUE_{intr} variation, and allow better choice of genotypes for these breeding strategies. Large variations in these traits were found, with variation in WUE_{intr} primarily being driven by $g_{H_2O}$ but also by considerable variation in photosynthetic capacity. Crucially, this independent variation did not lead to a trade-off; genotypes with moderately low $g_{H_2O}$ could have equally high photosynthetic rates due to high photosynthetic capacities. Finally, photosynthetic capacity advantages were maintained under all conditions, suggesting that breeding for this trait would increase WUE_{intr} under a broad spectrum of environments.

Supplementary data

Supplementary data are available at JXB online.

Figure S1. Rapid screening of the carboxylation capacity of Rubisco ($V_{cmax}$) and the maximal rate of RuBP regeneration ($J_{max}$) achieved for many replicates and genotypes as illustrated. Five points were measured at $C_s < 200 \mu$mol mol$^{-1}$ and the slope of these was taken as an estimate of the derivative of the classical Rubisco-limited photosynthesis function at a $C_i$ of $\Gamma^*$, the photo-compensation point. This estimate was used to solve for $V_{cmax}$ assuming an infinite internal conductance to CO$_2$ and the temperature corrections for Rubisco kinetic constants from von Caemmerer (2000). $J_{max}$ was estimated as four times the maximum rate of CO$_2$ and light-saturated photosynthesis at $>2000 \mu$mol mol$^{-1}$ CO$_2$. Thus, only one and five measurements were needed to provide high-throughput estimates of $J_{max}$ and $V_{cmax}$, allowing 178 and 91 leaves to be measured, respectively. Filled points represent the points used for estimation of $V_{cmax}$ and $J_{max}$, open points represent reference points measured for a full CO$_2$ response curve, but not used in the analysis.

Figure S2. Differences between genotypes in the relationship of maximum Rubisco carboxylation capacity ($V_{cmax}$) to leaf temperature for three genotypes with constant TR at high VPD (circles) and three genotypes with increasing TR at high VPD (triangles) under well-watered conditions. Each panel shows measurements made for a soybean genotype (filled symbols) and points for all genotypes combined (open squares). The fitted relationships (a power function) are shown for each genotype (solid line) and relative to the relationship for PI416937 (dashed line). Statistical differences and standard errors of the mean are shown in Table S2.

Figure S3. Differences between genotypes in the relationship of maximum RuBP regeneration capacity ($J_{max}$) to leaf temperature for all 11 genotypes under well-watered conditions. Each panel shows measurements made for a soybean genotype (filled symbols) and points for all genotypes combined (open squares). The fitted relationships (a linear function) are shown for each genotype (solid line) and relative to the relationship for PI416937 (dashed line). Statistical differences and standard errors of the mean are shown in Table S2. Soybean genotypes with a constant TR at high VPD (circles) and genotypes with increasing TR at high VPD (triangles) are shown.

Table S1. Minimum adequate linear models for stomatal conductance to H$_2$O ($g_{H_2O}$), and photosynthetic rate ($A$) measured during a drought experiment on 11 soybean genotypes.

Table S2. Minimum adequate linear model for the maximal rate of Rubisco carboxylation ($V_{cmax}$), and RuBP regeneration ($J_{max}$), measured on well-watered plants.

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